

EFFECT OF USING OF TWO PLANTS POWDER SILYBUM (*SILYBUM MARIANUM*) AND LICORICE (*GLYCYRRHIZA GLABRA*) ON TWO TYPES OF GROWTH LEGUMINOUS PLANTS IN SOILS POLLUTED BY HEAVY METALS

Wadullah Asaad Abdullah Al-mtewti and Hussein Saber Mohammed Ali Al-Rashedy

Department of Biology, College of Education for pure Science, University of Mosul, Mosul, Iraq.

Abstract

The research aims at studying the effect of two plants powder Silybum *Silybum marianum* and licorice *Glycyrrhiza glabra* by concentrations (0, 3, 6) g/kg soil for each of them on growth cowpea *Vigna unguiculata* and beans *Phaseolus vulgaris* plants in soils polluted by two elements of cobalt with concentrations (0, 30, 60) and copper with concentrations (0, 70, 100) mg/kg soil. The results showed that soil treatment with cobalt at concentrations (60) mg/kg soil led to significant increase by concentrating peroxidase enzyme where it totaled (0.095) μ m/mL, superoxide dismutase where it totaled (0.120) μ g/mL/min for the two plants compared by the control treatment, meanwhile the results showed that the soil treatment with powder licorice at concentration (3) g/kg soil led to insignificant decrease by concentrating superoxide enzyme dismutase for the two plants compared to the control treatment, but when the soil is treated by powder licorice at concentration (6) g/kg soil it was observed that it caused an insignificant decrease in the concentration of Peroxides enzyme for both plants compared to the control treatment.

Key words : Silybum marianum, Glycyrrhiza glabra, enzyme dismutase

Introduction

The increase in the proportion of the world's population has led to an increased interest in environmental pollution and poisoning by chemicals ,as the acceleration of industrial and urban processes led to the accumulation of pollutants and toxins such as pesticides, oil products, acids and heavy metals in natural resources such as soil , air and water Which led to a decrease not only in the quality of the environment, but also the impact on both animal and plant life. Heavy metals, including copper, cobalt, chromium, lead, cadmium, mercury and nickel, are important environmental pollutants that lead to plant poisoning and are a serious threat to various ecosystems (Sethy and Ghosh, 2013). Pollution of the environment with heavy metals comes from various sources, including industrial and agricultural sources, which are fields of tremendous interest in all parts of the world. Although heavy metals are natural components in the soil at very few concentrations, the high environmental pollutants resulting from industry, agriculture and mining operations have contributed to undesirable poisonous accumulations

of heavy metals (Chandra and Kang, 2016). Heavy metals such as cobalt, copper, iron, manganese, molybdenum, nickel and zinc enter the soil from various sources such as casting factories, mining, combustion and agricultural (Nagajyoti et al., 2010). The problem of pollution of ecosystem with heavy metals may also arise from the long-term use of untreated waste in irrigation, which led to an increase in the concentration of heavy metals in the soil (Lu et al., 2015). The heavy metals are considered important environmental pollutants and their toxicity is reckoned a grave problem where it arouses a huge concern for environmental, nutritional and toxicological reasons, as metals can affect a number of physiological and biochemical processes in the plants, and their toxicity varies with different types of plants, metal type, metal concentration and chemical form (Ackova, 2018). The soil is counted the fundamental part and most important in the environmental system where the soil is polluted significantly with heavy metals across world (Hakeem et al., 2014). And the areas polluted with heavy metal acquires a great interest because of its possible danger

on the safety and security of food and its harmful effects on the environmental system (Mombo *et al.*, 2015). Existence of heavy elements in the environment prevents plants from reaching their fullest potential in terms of growth and reproduction, because once they are deposited on the ground the plants are able to eat these elements from the soil and insert them into the food chain, which increases the risks of toxicity to metals on humans and animals (Roy and McDonald, 2015). Just as heavy metal pollution may severely hinder plant growth and productivity and it also increases the risk of harm to animal and human health through biomagnification (Sharma, 2012).

Materials and methods

Preparedness and soil treatment

The surface soil was taken from Governorate of Nineveh definitely area of Zayuna at depth of (0-30) cm and was air dried, then smoothened and passed through a sieve whose diameters of holes are (2) Mm. The treatments used in this study comprised two types of heavy elements namely cobalt and copper with two different concentrations for each element, and treating the soil with two types of powder of wild plants and they are Silybum Silybum marianum and licorice Glycyrrhiza glabra and in two concentrations each for them (the two plants' powder was obtained by taking the leaves of the plant for the Silybum and the roots of the licorice plant and drying them with air then they were ground using an electric mill to be ready for use in soil treatment, according to the type and concentration of the treatment), in addition to the control treatment and by three replications for each treatment and the elements were added ,each separately and powder of Silvbum plant and powder of licorice plant, each was added separately according to the treatment to soil, as it was mixed outside the pot and then placed in the pot to ensure that it blended well and the experiment was carried out using plastic pots whose diameter is (23) cm and height is (20) cm, capacity of each pot is (5) kg soil and the treatments were as follows:

A- Adding cobalt in the form of ($Co (No_3)_2.6H_2o$) to the pots soil with two concentrations (30, 60) mg/kg respectively.

B- Adding cobber in the form of ($\text{Cu} (\text{No}_3)_2.3\text{H}_2\text{O}$) to the pots soil with two concentrations (70, 10) mg/kg soil respectively.

C- The soil treatment with powder of Silybum with concentration of (3) and (6) g/kg respectively.

D- The soil treatment with powder of licorice with concentration of (3) and (6) g/kg respectively.

E- The interventions:

1- The soil treatment with powder of Silybum with concentration (3) g/kg + the soil treatment with cobalt with concentration (30, 60) mg/kg soil respectively.

2- The soil treatment with powder of Silybum with concentration (6) g/kg + the soil treatment with cobalt with concentration (30, 60) mg/kg soil respectively.

3- The soil treatment with powder of Silybum with concentration (3) g/kg + the soil treatment with copper with concentration (70, 100) mg/kg soil respectively.

4- The soil treatment with powder of Silybum with concentration (6) g/kg + the soil treatment with cobber with concentration (70, 100) mg/kg soil respectively.

5- The soil treatment with powder of licorice with concentration (3) g/kg + the soil treatment with cobalt with concentration (30, 60) mg/kg soil respectively.

6- The soil treatment with powder of licorice with concentration (6) g/kg + the soil treatment with cobalt with concentration (30, 60) mg/kg soil respectively.

7- The soil treatment with powder of licorice with concentration (3) g/kg + the soil treatment with copper with concentration (70, 100) mg/kg soil respectively.

8- The soil treatment with powder of licorice with concentration (6) g/kg + the soil treatment with copper with concentration (70, 100) mg/kg soil respectively.

Agriculture and Irrigation

Cowpea Vigna unguiculata and beans Phaseolus vulgaris were obtained at shops for selling seeds and agricultural items in the city of Mosul in Iraq. The seeds were planted on 21/8/2019 (15) seeds/pot and it was taken into consideration that the distances should be equal between seeds, the pots were placed randomly under circumstances of plastic house and after passage of (60) days of agriculture date the cowpea and beans were harvested with (3) three replications per treatment.

The studied characteristics

Peroxidase enzyme effectiveness estimate

The peroxidase enzyme effectiveness was measured in the plant leaves and absorbance was read using the Spectrophotometer at wave length (470) nm according to method mentioned in (Badea *et al.*, 1999) the following law was applied :

$$A = \pounds \times L \times C$$
$$C = A / 26.6$$

A = The absorbance

 $C = Enzyme \text{ effectiveness } (\mu m/mL)$

f = Completion coefficient equals (26.6)

JUAIL AIL	Effect	ofplant	type			0.055	q	0.066	а																				
	/kg	orice	9	090.0	lid	0.054	b-u	0.066	.R																				
naint	00 mg	Lic	3	0.069	de	0.068	h-k	0.069	g-k																				
Ind IIn	Cu 1	unc	9	0.068	def	0.065	jkl	0.070	g-k																				
		Silyt	e	0.065	efg	0.055	dou	0.075	fg											0.067	A	0.072	A	<u>il (5%</u>					
BIUWI		orice	9	0.070	q	0.082	e	0.057	no									type						ty leve					
cillarid	mg/k	Lic	e	0.064	fg	0.059	oum	0.069	<u>ъ</u> ч									plant 1						babili					
cilip	Cu 70	unc	9	0.062	ghi	0.059	mno	0.064	klm									The						ne pro	est				
מ מווח	ľ	Silyl	e	0.063	G	0.053	bdo	0.073	Ē									type ×						ly at tl	nialy t				
cow he		rice	9	0.057	. –	0.058	no	0.056	no									ement						ificant	olynon				
	g CO	Lico	e	0.069	de	0.070	g-k	0.067	ijk									t of ele						r sign	can po				
	l/gm (m	و	0.059	÷Ēŕ	0.044	st	0.073	£									Effec						t diffe	e Dun				
	90	Silyb	e	0.065	efg	0.055	dou	0.074	fgh											0.058	a	0.071	a	s don	to the				
cuu ai		rice	9	0.053 (k	0.037 (٩v	0.068 (h-k											Ŭ		•		r letter	ording				
	g CO	Lico	e) 990.(d-g	090.0	lmn	0.071	£.															imila	acci				
CENTR	mg/k	mn	9	0.075 (· 	0.038 (٧N	0.076	f															with s					
n per c	30	Silyb	e	0.042 (-	0.033 (WW	0.050 (pqr															tions					
		<u>5</u>	100	0.088 (р	0.087 (b-e) 680.(\mathbf{bc}	It		70		ıt		0.070	a	tion	Se	990.(a	0.074	a	equa					
ý how	J	mg/]	70	0.086 (\mathbf{bc}	0.083 (de	0.088 (bcd	elemen	ē	0.0	а	elemei	ration	0690.0	a	centra	ant typ	0.067 (a	0.070 (а	The					
וורטוור		kg	60	095 (A) 160.	A	092 (Ab	ct of e	typ	54		ct of e	ncenti) 690;	A	nt con	The pl	065 (A	0.072	A						
וו מוות		mg/	30	0.084 (с	080	cde	0.082 (e	Effe		0.0	A	Effe	00	090.0	a	eleme	×	0.051 0	a) [690]	а						
IIJUUI	ice	6.0	9	034 (u	018 (x	049 (qrs			.055	а	88		055 (a			020 (a	029 (а)e		71		62	
	Licor	g/k	e	038 0	lmn	0.031 0	M	045 0	rst			0.061 0	a	0.05	а	0.061	a	ion ×		0.058 0	a	0.064 0	a	ant ty		2.6	а	0.0	а
	mn	5.0	9	0.036 (um	0.028 (M	0.044 (st	0.056	a			55		0.056 (а	entrat	ē	0.047 (a	0.065 (a	The pl		1 6		4	
וו נו כמנ	Silyb	g/k	e	0.035 (um	0.028 (M	0.041 (tu	0.054 (а			0.0	а	0.054 (a	r conc	ant typ	0.045 (a	0.063 (а	pe × J		0.0	а	0.05	а
pper.	ontrol		1	0.035	um	020	x	.049	qrs	silybu	tration	licorice	tion	ļ	pe	owde	ation	of powde	The pl					powder ty					
	The C	lant	vpe) -wo	ea	seans (ffect of S	1 concen	ffect of 1	oncentra	ffect of	owder ty	ffect of p	concenti	Effect		Jowpea		seans		ffect of p		Jowpea		seans	
21		d	÷.			\mathbf{C}	þ	Ш		ш	Ц	ш	ပ	щ	ф	Ш	r			\cup		щ		ш				щ	

um /mL) for cownea and heans plants growth in soil polluted with cobalt and concentration(**Table 1:** Effect of soil treatment with Silvhum and licorice nowder on neroxidase

with	cobalt and c	opper	·																				
The Cont	rol Silyb	um	Licorid	ce	CO		Cu	3() mg/	kg C(<u> </u>	60	mg/k	g CO		Ü	u 70 n	ıg/kg		Ú	u 100	mg/kg	Effec
plant	g/k	0,c	g/kg		mg/kg	В	g/kg	Sily	mnq	Lico	orice	Silyb	m	Licol	rice	Silybı	m	Licor	ice S	Silybu	Е	Licoric	e of pla
type	e	9	3 9 9	- -	30 60	70	100	e	9	e	9	e	9	e	9	e	9	e	9	e	9	3 6	type
0.03	4 0.047	0.045 (0.033 0.0)48 <u>0</u>	.112 0.12	<u>30 0.09</u>	3 0.11	8 0.048	0.083	0.064	0.074	0.063 ().100 () 0.07) <u>.095 (</u>	0.051 0	0.078 0	077 0	0.075 0	065 0.	078 0.	95 0.09	9
ш	1	-	В	-	b A	e	а	-	f	÷Ē	q	. –	c	· 	de	k	ac	G	gh	ij	ac	d d	
Cow- 0.02	0.0 0	0.048 (0.048 0.0) <u>50 0.</u>	.153 0.16	6 0.11	6 0.13	1 0.054	0.112	0.078	0.093	0.084 ().129 () 080 ().129 (0.056 0	0.110 0	088 0	0.105 0	067 0.	110 0.	99 0.10	6 0.092
pea x	st	tuv	tuv s1	tu	b A	q	q	rs	de	-	• =	Ч	c	k	c	r	ef	jk	ы ц	dou	ef	h fg	а
Beans 0.04	8 0.041	0.041 (0.018 0.0	<u>145 0.</u>	071 0.07	73 0.06	9 0.10	5 0.041	0.0540	050	0.054	0.041 (0.071 (0.048 (0.061 (0.046 0	0.046 0	065 0	045 0	063 0.0	046 0.	91 0.08	6 0.057
tuv	w	Μ	x	N N	mn M	m	ы 0	Ŵ	rs	stu	IS	Μ	um	tuv	Ь	uv	nv (pqC	M	bd r	١٧	ij k	q
Effect of Sily	bu 0.055 (0.077	-		Ë	ffect of	4.													-		-	-
m concentrat	ion b	а			elen	nent ty	be																
Effect of Lic	orice		0.066 0.0	111	0.083		.082																
concentratio	u		b ŝ	г	A		a																
Effect of	0.0	67	0.072		Effect	of elen	nent	1															
powder type	а		q		conc	entrati	on																
Effect of pow	der 0.055 (0.077 (0.066 0.0	0.77	0.06 0.08	<u>39 0.07</u>	5 0.08	6															
concentratio	u c	а	b ŝ	7	b A	q	а																
Effect of pow	der concen	tration	1 ×	e	lement (concer	ntration					Eff	set of 6	slemer	it type	× Th	e plant	type					
The plant ty	ЭС				\times The	plant	type																
Cowpea	0.063	0.102 (0.078 0.0	97 0	098 0.11	<u>60.0 6</u>	5 0.10				0.108								0	860			
	ပ	а	b 3	7	b A	q	q				а									A			
Beans	0.046	0.052 (0.054 0.0)58 <u>0</u> .	054 0.05	50 0:02	4 0.07	~			0.056								0	066			
	q	cd	cd c	p	d D	q	ပ				а									A			
Effect of pov	der type ×]	The pl	ant type				H	he equi	ations	with :	similar	- letter	s don't	differ	signif	icantly	/ at the	proba	bility	level ((2%)		
											acco	ording	to the	Dunc	an pol	ynomi	aly tes	Ļ,					
Cowpea	0.092	-	0.087																				
	а		а																				
Beans	0.049 h	-	0.056 h																				

Table 2: Effect of soil treatment with Silybum and licorice powder on superoxide dismutase concentration (µg/mL/min) for cowpea and beans plants growth in soil polluted

Г

L = Cell thickness equals (1)

Superoxide dismutase enzyme (SOD) effectiveness estimate

SOD enzyme effectiveness (unit/1g fresh weight) was measured in plant leaves and the absorbance was read using the Spectrophotometer at wave length (480) nm according to method (Bosco *et al.*, 2007)mentioned in (Misra and Fridovich, 1972).

Statistical Analysis

The experiments were designed and statistically analyzed using the practical experiment according to completely randomized design (C.R.D) in the factorial experiments (Antar, 2010), the comparison was completed between the significant differences in the treatments rates using (Duncan's new multiple range test).

Results and Discussion

Peroxidase Enzyme (POD) Estimate

The table 1 states that the soil treatment with licorice Glycyrrhiza glabra powder at concentration (6) g/kg soil led to decrease occurrence by concentration of peroxidase enzyme in the leaves tissues of the two plants compared to the treatment of control and other treatments where the decrease totaled $(0.034) \,\mu\text{m/mL}$. As noted that the soil treatment with the element of cobalt at concentration (60) mg/kg led to occurrence of significant increase by concentrating peroxidase enzyme in the leaves tissues of both cowpea and beans plants where the increase totaled (0.095) µm/mL compared to the control treatment and other treatments . Also the table 1 stated that silvbum Silvbum marianum powder addition at concentration (3) g/kg soil to soils polluted with cobalt at concentration (30) mg/kg soil led to decease of concentration of peroxidase enzyme in the leaves tissues of the two plants compared to growth plants with soils polluted with heavy elements and soils polluted with heavy elements and treatment with plant powders where it totaled (0.042) µm/mL. Increase in concentration POD of cowpea and beans plants at treatment with heavy metals may be attributed to increase of levels of hydrogen peroxide in the plants stressed with heavy metal and the changes in POD activity in the plants cells are considered vital and good indicators for the vital and non-vital stresses (Doganlar and Atmaca, 2011). But the cause of decrease in POD concentration of cowpea and beans plants at treatment with licorice powder may be attributed to existence of metals nutrients in licorice extract, it has increased the plant's tolerance to heavy metals by preserving the mechanisms of photosynthesis and reducing the impact of heavy metals on the cell

membrane, leading to a decrease or reduction in the concentration of antioxidant enzymes, including POD (Bargaz *et al.*, 2016).

Estimate of superoxide dismutase Enzyme (SOD)

The table 2 states that the soil treatment with licorice Glycyrrhiza glabra powder at concentration (3) g/kg soil led to significant decrease at concentration SOD for the two plants compared to the other treatments where it totaled (0.033) µg/mL/min with exception of insignificant decrease with the control treatment. As observed that the soil treatment with cobalt element at concentration (60) mg/kg led to significant increase at concentration SOD enzyme in the leaves tissues for both beans and cowpea where it totaled $(0.120) \mu g/mL/min$ compared to the control treatment and other treatments except insignificant superiority at copper treatment at concentration (100) mg/kg soil. Also, the table 2 states that addition of Silybum Silybum marianum powder rat concentration (3) g/kg soil to the soils polluted with cobalt at concentration (30) mg/kg soil led to significant decrease at concentration SOD enzyme in the leaves tissues for the two plants compared to growth plants with soils polluted with heavy elements and soils polluted with heavy elements treatment with plants powders where it totaled (0.048) µg/mL/min. The reason for the increase in concentration SOD plants for cowpea and beans plants when dealing with heavy metals may be due to the high concentrations of antioxidant enzymes that have entered into the protection mechanisms adopted by plants, which is associated with increasing the mitigation against high stress (Awasthi and Sinha, 2013). For the decrease in the concentration of SOD plants for cowpea and beans when dealing licorice powder may be due to the fact that licorice extract contains soluble sugars, which have an important physiological role In supplying the plant with energy and preserving the water potential of leaves and plant water content, which reduces the cellular osmotic pressure and increases the tolerance of heavy metals, which leads to reduce the damage of stress on the cell membrane and thus decrease the concentration of some antioxidant enzymes including SOD (Babaeian et al., 2011).

References

- Ackova, D.G. (2018). Heavy metals and their general toxicity on plants. *Macedonia Plant Science Today*, **5(1):** 14-18.
- Antar, S.H. (2010). Statistical analysis in scientific research and SAS program. College of agriculture and Forestry, University of Mosul, Dar Ibn Al-Atheer house for Printing and publishing, Mousl, Iraq. (Arabic).
- Awasthi, K. and P. Sinha (2013). Nickel stress induced

antioxidant defense system in sponge gourd (*Luffa cylindrica*). Journal of Plant Physiology and Pathology, **1**: 1.

- Babaeian, M., A. Tavassoli, A. Ghanbari, Y. Esmaeilian and M. Fahimifard (2011). Effects of foliar micronutrient application on osmotic adjustments, grain yield and yield components in sunflower (*Alstar cultivar*) under water stress at three stages. *African Journal of Agricultural Research*, 6(5): pp. 1204-1208.
- Badea, E., G. Marinescu, C. Babeansu, M. Corneanu and G. Corneanu (1999). Peroxidase Isoenzyme Pattern and total activity in *Arachis hypogaea* In vivo culture under conditions similar to Extraterrestrial environment. *Plant Peroxidase Newsletter*, 14: 139-146.
- Bargaz, A., M.A. Rania, S.G. Maybelle and R. Mostafa (2016). Improved salinity tolerance by phosphorus fertilizer in two *Phaseolus vulgaris* recombinant inbred lines contrasting in their P-efficiency. *Journal of Agronomy and Crop Science*, 202(6): 497-507.
- Bosco, G., Zh. Yang, J. Nandi, J. Wang, Ch. Chen and E.M. Comporesi (2007). Effect of hyperbaric oxygen on glucose, lactate, glycerol and anti -oxidant enzymes ischemia and reperfusion. *Clinical and Experimental Pharmacology* and Physiology, 34(1-2): 70-76.
- Chandra, R. and H. Kang (2016). Mixed heavy metal stress on photosynthesis, transpiration rate and chlorophyll content in poplar hybrids, forest science and technology, *Forest Science and Technology*, **12(2)**: 55-61.
- Doganlar, Z.B. and M. Atmaca (2011). Influence of airborne pollution on Cd, Zn, Pb, Cu and Al accumulation and

physiological parameters of plant leaves in Antakya (Turkey). *Water Air Soil Pollut.*, **214:** 509-523.

- Hakeem, K.R., M. Sabir, M. Ozturk and A. Mermut (2014). Soil remediation and plants: prospects and challenges. Elsevier, Waltham, pp 1-724.
- Lu, Y., H. Yao, D. Shan, Y. Jiang, S. Zhang and J. Yang (2015). Heavy metal residues in soil and accumulation in maize at long term wastewater irrigation area in Tongliao, *China. J. Chem.*, 2015: 628280.
- Misra, H.P. and I. Fridovich (1972). The role of Superoxide anion in the auto-oxidation of epinephrine and a simple assay for SOD. *J. Biol. Chem.*, **247**: 317.
- Mombo, S., Y. Foucault, F. Deola, I. Gaillard, S. Goix, M. Shahid, E. Schreck, A. Pierart and C. Dumat (2015). Management of human health risk in the context of kitchen gardens polluted by lead and cadmium near a lead recycling company. J. Soils Sediments, doi: 10.1007 / s11368-015-1069-7.
- Nagajyoti, P.C., K.D. Lee and T.V.M. Sreekanth (2010). Heavy metals, occurrence and toxicity for plants: *a review*. *Environ. Chem. Lett.*, 8: 199-216. doi: 10.1007/s10311-010-0297-8.
- Roy, M. and L.M. McDonald (2015). Metal uptake in plants and health risk assessments in metal-contaminated smelter soils. *L. Degrad Dev.*, 26: 785-792.
- Sethy, S.K. and S. Ghosh (2013). Effect of heavy metals on germination of seeds. *Journal of Natural Science, Biology and Medicine*, **4(2):**.
- Sharma, I. (2012). Arsenic induced oxidative stress in plants. *Biologia*, **67(3):** 447-453.